



Visual Restoration And Circuitry -

Retinal Sheet Transplants To Rats With Retinal Degeneration



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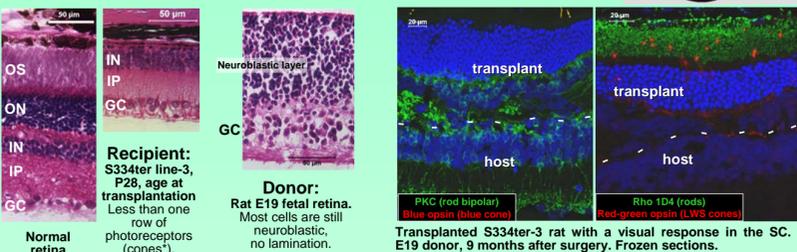
PURPOSE

To investigate which cell types are involved in the transplant-induced visual restoration of a degenerating retina.

Background

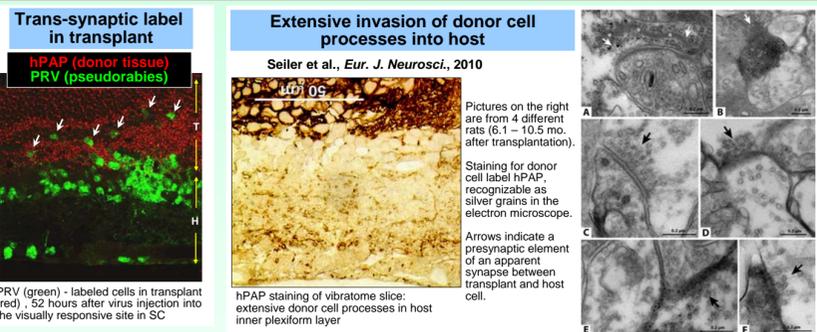
Transplantation of retinal sheets

Sheets of retinal progenitor cells, transplanted to the subretinal space of different rodent degeneration models, develop lamination like a normal retina and restore function.



Restoration of visual responses by transplants involves synaptic connectivity:

- Transplants preserve optokinetic responses (Thomas et al. 2004, *J Neurosci Meth*, 138:7-13)
- Light stimulation of the eyes can be recorded in the brain according to the retinotopic map (e.g., Seiler et al. 2008, *Eur J. Neurosci*, 28:208-220; Yang et al. 2010, *Exp. Eye Res.* 91: 727-738)
- Transplant neuronal cells can be labeled from the visually responsive site in the brain by trans-synaptic tracing (using labeled neurotropic pseudorabies virus) (Seiler et al., *Eur. J. Neurosci* 2005, 2008)
- Transplant processes invade the host retina and form synapses in the host inner plexiform layer (Seiler et al., *Eur J. Neurosci.* 2010)



Background summary: Which specific cell types are involved in the circuitry between transplant and host retina?

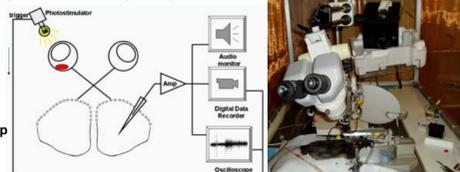
Methods

1. Transplantation

S334ter-line-3 rats (age 0.8 - 1.4 months) with fast retinal degeneration received E19 fetal retinal sheet transplants derived from transgenic rats expressing hPAP (human placental alkaline phosphatase). Some donor retinas were coated with BDNF microspheres or GDNF microspheres before implantation (Seiler et al. 2008b). Rat eyes were imaged by 3D-Ocular Coherence Tomography (OCT) 0.4 - 2.2 months after surgery to determine transplant placement and layering (Seiler et al. 2010b). Thirteen transplanted rats were selected and sacrificed at 2.3 - 8.5 months after transplantation (age 3.4 - 9.9 months). Eight of these rats were recorded by electrophysiology for visual responses in the superior colliculus (below).

2. SC recording

Multiunit visual responses were recorded from the exposed superior colliculus (SC) with a 40ms full-field illumination stimulus varying from -5.9 to 1 log cd/m² (see Yang et al. 2010, *Exp. Eye Res.* 91:727-738).



3. Tissue processing

Rats were perfused through the heart into the ascending aorta with 2.5% glutaraldehyde, 1% paraformaldehyde, 3% sucrose, 1mM MgSO₄ in 0.1M phosphate buffer. Eye cups were postfixed overnight after removal of the cornea and embedded in 4% agarose for vibratome sectioning. Selected vibratome slices were flat embedded in Eponate.

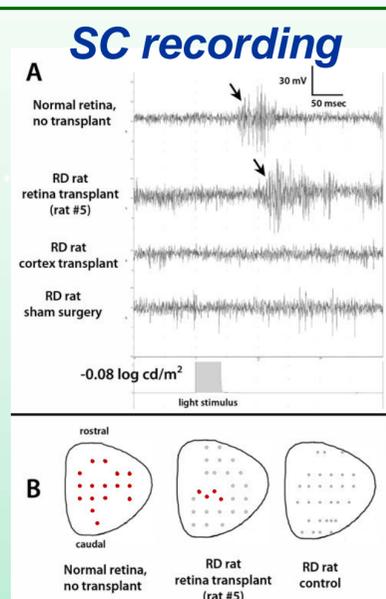
4. Immunohistochemistry for Molecular Phenotyping

Blocks were thin sectioned at 0.25 μm into serial arrays and probed for aspartate, glutamate, glycine, glutathione, glutamine, arginine, taurine, GABA, rhodopsin, cone opsin, CRALBP, and DAPI (procedure after Jones et al. 2003 (*Comp. Neurol.* 464:1-16). http://prometheus.med.utah.edu/~marclab/protocols_CMP_Html

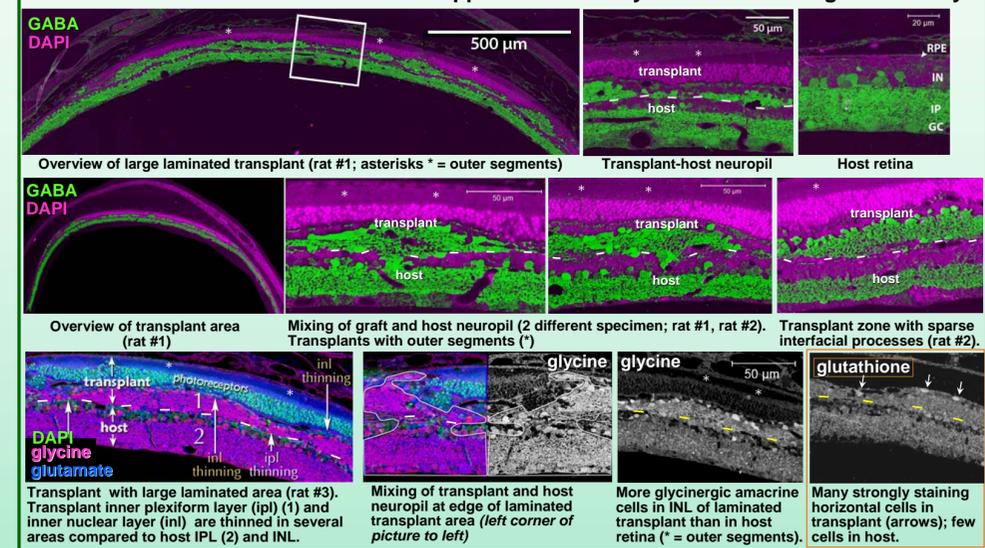
RESULTS

1 Overview of experiments

Rat #	Sex	Donor tissue	Age at surgery (mo)	Survival time (mo)	Age at sacrifice (mo)	Organization	Remarks
1	F	E19 retina	1.3	8.5	9.8	Laminated area (7 of 12 slices)	not recorded
2	M	E19 retina + BDNF	1.4	8.3	9.7	Laminated area (6 of 12 slices)	not recorded
3	F	E19 retina + BDNF	1.2	8.4	9.6	Laminated area (3 of 11 slices)	not recorded
4	F	E19 retina + GDNF	0.8	6.7	7.5	Mostly rosettes, laminated area in center of graft (4 of 9 slices)	not recorded
5	F	E19 retina + GDNF	1.1	2.9	4.0	Mostly rosettes, small laminated area (5 of 11 slices)	SC recording, threshold -2.7 log cd/m ²
6	F	E19 retina + BDNF	1.2	8.4	9.6	Laminated area (7 of 11 slices)	not recorded
7	F	E19 retina + GDNF	0.9	4.3	5.2	Laminated area (9 of 13 slices)	SC recording, threshold -2.2 log cd/m ²
8	F	E19 retina + GDNF	1.1	4.3	5.4	Laminated area (5 of 12 slices)	SC recording, threshold -1.5 log cd/m ²
9	F	E19 retina + GDNF	1.1	2.6	3.7	Laminated areas (8 of 11 slices)	SC recording, threshold -2.0 log cd/m ²
10	M	E19 retina	1.1	2.9	4.0	Mostly rosettes, small laminated area (3 of 10 slices)	SC recording, threshold -2.5 log cd/m ²
11	F	E19 retina + BDNF	1.2	3.1	4.3	Mainly rosettes, tiny laminated area (5 of 12 slices)	SC recording, threshold -2.2 log cd/m ²
12	F	E19 retina + GDNF	0.8	4.2	5.0	Rosettes only (7 slices)	SC recording, threshold -0.9 log cd/m ²
13	M	E19 retina + GDNF	1.1	2.3	3.4	Rosettes only (11 slices)	SC recording, threshold -1.5 log cd/m ²

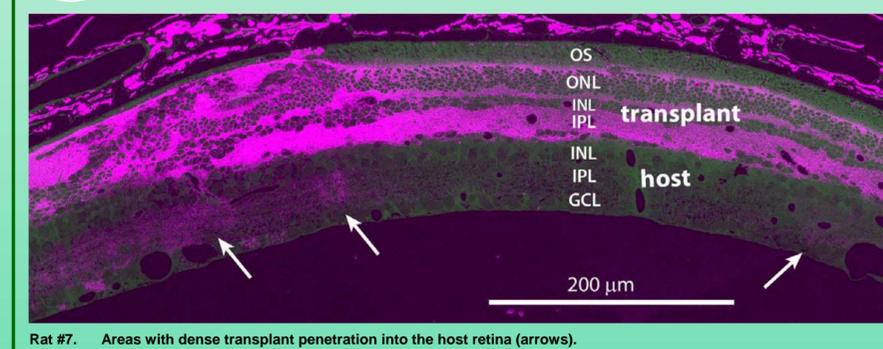


4 Inner retinal neurons (GABA, glutamate, glutathione)

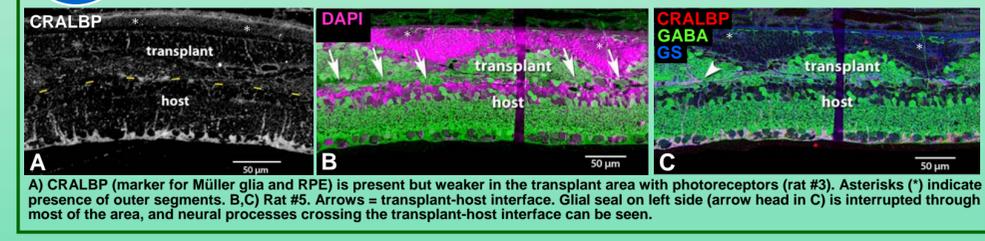


2 Donor cell label (hPAP)

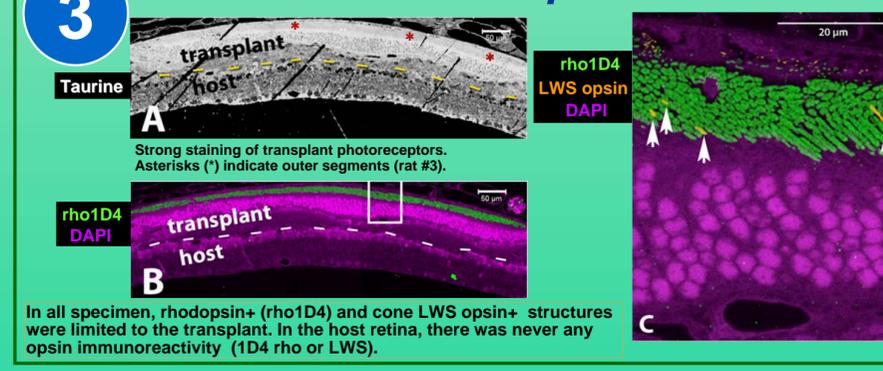
The donor tissue was derived from transgenic rats expressing human placental alkaline phosphatase (hPAP). hPAP was detected histochemically on vibratome slices with BCIP/NBT (Sigma). Slices were embedded in Eponate and sectioned. Magenta = NBT (transplant signal); green = autofluorescence.



5 Glial markers (glutamine synthetase, CRALBP) & neuronal marker (GABA)



3 Photoreceptors



CONCLUSIONS

- The host photoreceptors are degenerated. Transplants contain healthy rods & cones with outer segments.
- The data indicate that amacrine and horizontal cells of donor and host are mainly involved in the function and communication between transplant and host.
- An extensive loss of bipolar cells was observed in the host and to a lesser extent in transplant.
- Host and graft neuropil are mixed in many areas, with no glial barriers between transplant and host.

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Abbreviations:
H - Host
T - Transplant
GC - Ganglion cell layer
IP - inner plexiform layer
IN - inner nuclear layer
ON - outer nuclear layer
OS - outer segments
RPE - retinal pigment epithelium
hPAP - Human placental alkaline phosphatase (donor marker)
BCIP - substrate for alkaline phosphatase
LWS - long wavelength (red-green)
CRALBP - cellular retinaldehyde binding protein
GS - glutamine synthetase

References:
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